

Antigen Affinity Purification of Antibodies Protocol

Cytokines are signaling proteins necessary for cell-to-cell communication throughout the body. In accordance with their role in preserving vital biological functions, the amino acid sequence of homologous cytokines from different mammalian species is generally highly conserved, rendering them poor immunogens for antibody production.

Consequently, the sera obtained from host animals after immunization with cytokines (Figure I-1) typically contain minute amounts of cytokine-specific antibodies. Isolation of these antibodies by standard purification procedures (e.g. ion-exchange chromatography) is tedious and ineffective. On the other hand, purification protocols which exploit the binding affinity of antibodies to certain biomolecules have yielded much better results.

For example, Protein A and Protein G are bacterial (*Staphylococcus*)-derived proteins that possess high binding affinity toward the Fc

(Fragment crystallizable) region of immunoglobulins (IgGs), and when attached to a solid-support matrix, enable efficient separation of IgGs from other serum constituents. Protein A/G affinity purification of serum-IgGs typically enriches the desired antibody by more than 100-fold. Yet, in the case of anti-cytokine antibodies, the total IgG fraction often contains less than 0.2% of the desired antibody.

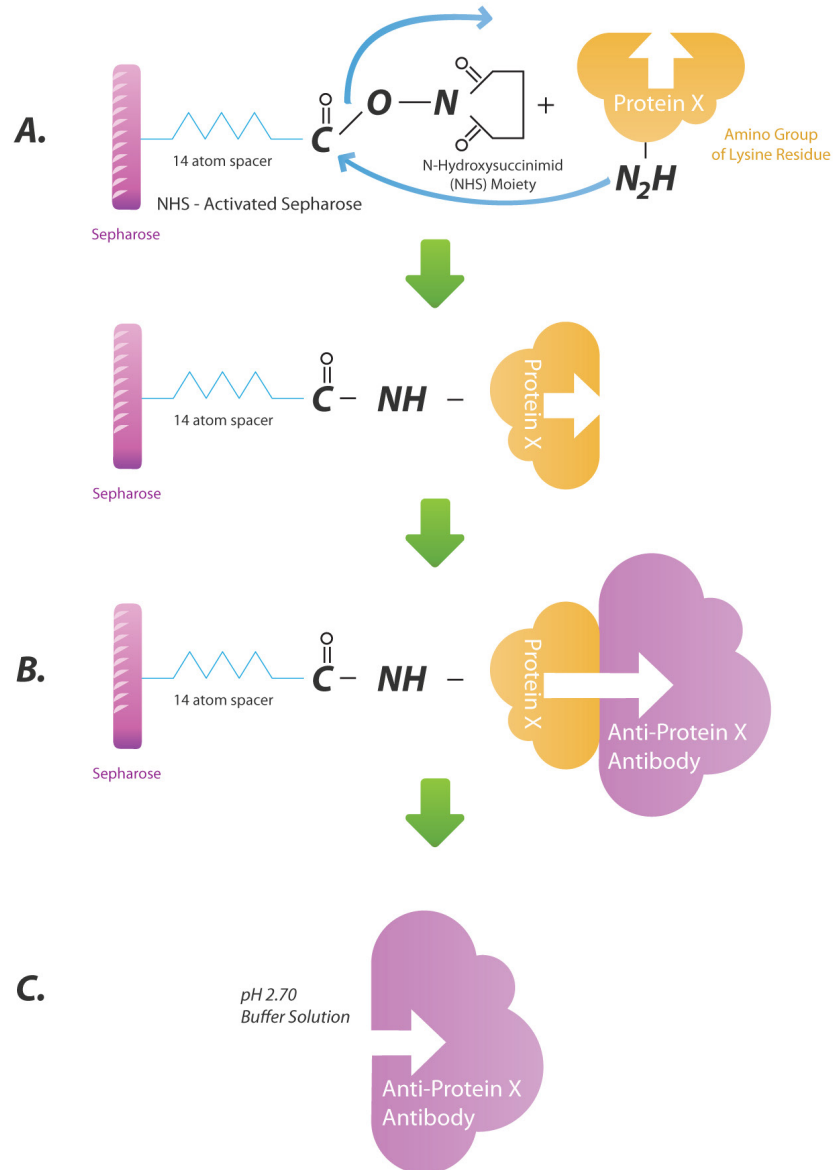
The large quantity of unrelated IgGs found in these preparations not only interferes with quantification of the relative amount of the cytokine-specific antibody, but also considerably increases the background noise when the antibody is used in analytical procedures such as ELISA, neutralization, immunohistochemistry, and Western blotting. A superior method for isolating specific polyclonal antibodies from antiserum is by affinity chromatography, which exploits the specificity of antibody-antigen interactions (Figure I-2).

Figure I-1 - General Immunization Schedule for Polyclonal Antibody Production in Rabbits

Day	Procedure	Antiserum Collected
1	Prebleed	3ml
1	Initial Immunization (250ug)	-
21	Immunization (125ug)	-
31	Titer Test Bleed	3ml
42	Immunization (125ug)	-
49	Production Bleed	10-25ml
56	Production Bleed	10-25ml
63	Immunization (125ug)	-
70	Production Bleed	10-25ml
77	Production Bleed	10-25ml
84	Immunization (125ug)	-
91	Production Bleed	10-25ml
98	Production Bleed	10-25ml

Figure I-2 -Antigen Affinity Purification of Antibodies

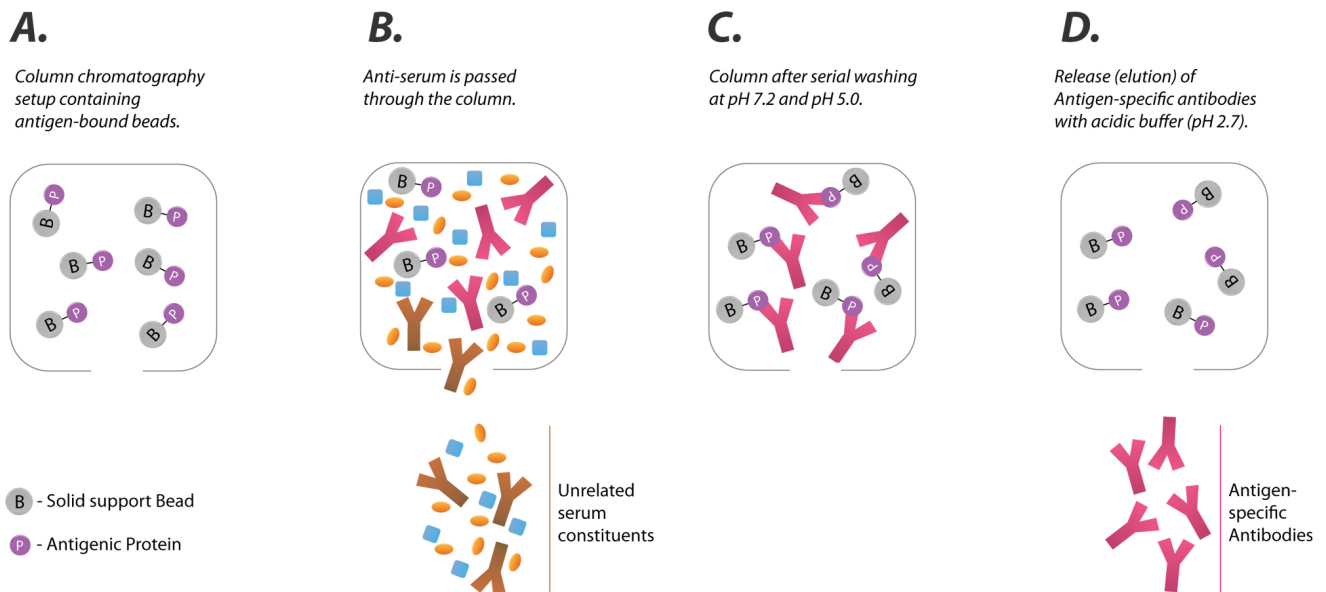
(A.) Preparation of separation resin.
(B.) Specific binding of Antibody to the resin
(C.) pH elution of the antibody from the affinity column.



Antigen Affinity Purification of Antibodies Protocol continued...

Here, the separation medium consists of a solid-support resin to which the antigen is attached through stable covalent bonds. The immobilized antigen is then used in a column chromatography setup to selectively capture antigen-specific antibodies, while other serum proteins and unrelated immunoglobulins are washed away (Figure I-3).

Figure I-3 - Schematic illustration of column chromatography setup for antigen affinity purification of antibodies.



NOTE: Elution is titrated with 1M NaOH to pH 7.2.

Antigen Affinity Purification of Antibodies Protocol continued...

The antigen-bound antibodies can be eluted from the column by an acidic solution (Figure I-4), which is promptly brought back to physiological pH to prevent acid-catalyzed antibody denaturation. This method typically yields >95% pure specific antibodies (Figure I-5).

Figure I-4 -
A Typical Antigen Affinity Purification Chromatogram

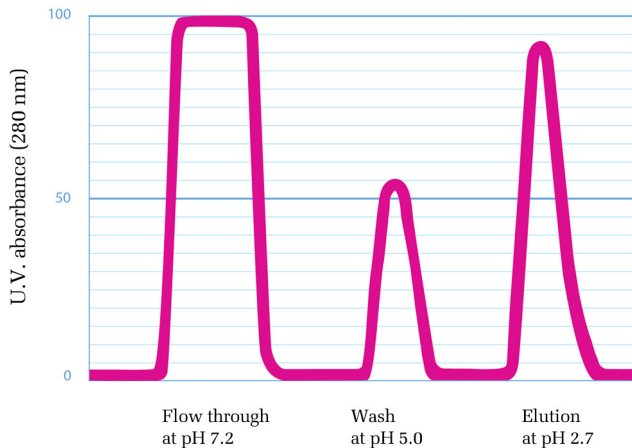


Figure I-5 -
SDS-PAGE analysis of Rabbit Anti-Human IL-8 polyclonal antibodies (RbahIL-8)

Lane 1: Molecular weight markers
Lane 2: 2 μ g of RbahIL-8 (unreduced)
Lane 3: 2 μ g of RbahIL-8 (reduced)

